

EXHIBIT B

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A program for construction of a manufactured IFN-Con DNA sequence was developed and is set out below in Table VIII. In the table, an asterisk designates the variations in IFN- α F needed to develop LeIFN-Con1, i.e., to develop the [Arg²², Ala⁷⁶, Asp⁷⁸, Glu⁷⁹, Tyr⁸⁶, Tyr⁹⁰, Leu⁹⁶, Thr¹⁵⁶, Asn¹⁵⁷, Leu¹⁵⁸] analog of IFN- α F. The illustrated top strand sequence includes, wherever possible, codons noted to the subject of preferential expression in *E. coli*. The sequence also includes bases providing recognition sites for Sal, HindIII, and BstE2 at positions intermediate the sequence and for XbaI and BamHI at its ends. The latter sites are selected for use in incorporation of the sequence in a pBR322 vector, as was the case with the sequence developed for IFN- α F and its analogs.

-1 1 10
 Met Cys Asp Leu Pro Gln Thr His Ser Leu Gly Asn Arg Arg Ala
 ATG TGT GAT TTA CCT CAA ACT CAT TCT CTG GGT AAC CGT CCG CTT
 20
 Leu De Leu Leu Ala Gln Met Arg Arg Ile Ser Pro Phe Ser Cys
 CTG ATT CTG CTG GCA CAG ATG CGT CCG ATT TCC CCG TTT AGC TGC
 30
 Leu Lys Asp Arg His Asp Phe Gly Phe Pro Glu Glu Glu Phe Asp
 CTG AAA GAC CGT CAC GAC TTC GGC TTT CCG CAA GAA GAG TTC GAT
 40
 50
 Gly Asp Gln Phe Glu Lys Ala Glu Ala Ile Ser Val Leu His Glu
 GGC AAC CAA TTC CAG AAA AAT CAG GCA ATC TCT GTA CTG CAC GAA

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Example 7 which had been altered to specify threonine and methionine as residues 14 and 16, respectively. More specifically, [Thr¹⁴, Met¹⁶] IFN- α F, designated IFN- α F₂, was expressed in *E. coli* upon transformation with a vector of Example 7 which had been cut with 5 SalI and HindIII and into which a modified subunit II (of Table VII) was inserted. The specific modifications of subunit II involved assembly with segment 39 altered to replace the alanine-specifying codon, GCT, with a threonine-specifying ACT codon and replace the 10 isoleucine-specifying codon, ATT, with an ATG codon. Corresponding changes in complementary bases were made in section 40 of subunit LeuIFN-FII.

The following Example 9 and 10 relate to practice of the invention in the microbial synthesis of consensus 15 human leukocyte interferon polypeptides which can be designated as analogs of human leukocyte interferon subtype F.

EXAMPLE 9

"Consensus human leukocyte interferon" ("IFN-Con," "LeuIFN-Con") as employed herein shall mean a non-naturally-occurring polypeptide which predominantly includes those amino acid residues which are common to all naturally-occurring human leukocyte 25 interferon subtype sequences and which includes, at one or more of those positions wherein there is no amino acid common to all subtypes, an amino acid which predominantly occurs at that position and in no event includes any amino acid residue which is not extant in that position in at least one naturally-occurring subtype. (For purposes of this definition, subtype A is positionally aligned with other subtypes and thus reveals a "missing" amino acid at position 44.) As so defined, a consensus human leukocyte interferon will ordinarily 30 include all known common amino acid residues of all subtypes. It will be understood that the state of knowledge concerning naturally-occurring subtype sequences is continuously developing. New subtypes may be discovered which may destroy the "commonality" of 40 a particular residue at a particular position. Polypeptides whose structures are predicted on the basis of a later-amended determination of commonality at one or more positions would remain within the definition because they would nonetheless predominantly include common 45 amino acids and because those amino acids no longer held to be common would nonetheless quite

likely represent the predominant amino acid at the given positions. Failure of a polypeptide to include either a common or predominant amino acid at any given position would not remove the molecule from the definition so long as the residue at the position occurred in at least one subtype. Polypeptides lacking one or more internal or terminal residues of consensus human leukocyte interferon or including internal or terminal residues having no counterpart in any subtype would be considered analogs of human consensus leukocyte interferon.

Published predicted amino acid sequences for eight cDNA-derived human leukocyte interferon subtypes were analyzed in the context of the identities of amino acids within the sequence of 166 residues. See, generally, Goeddel, et al., *Nature*, 290, pp. 20-26 (1981) comparing LeIFN-A through LeIFN-H and noting that only 79 amino acids appear in identical positions in all eight interferon forms and 99 amino acids appear in identical positions if the E subtype (deduced from a cDNA pseudogene) was ignored. Each of the remaining positions was analyzed for the relative frequency of occurrence of a given amino acid and, where a given amino acid appeared at the same position in at least five of the eight forms, it was designated as the predominant amino acid for that position. A "consensus" polypeptide sequence of 166 amino acids was plotted out and compared back to the eight individual sequences, resulting in the determination that LeIFN-F required few modifications from its "naturally-occurring" form to comply with the consensus sequence.

A program for construction of a manufactured IFN-Con DNA sequence was developed and is set out below in Table VIII. In the table, an asterisk designates the variations in IFN- α F needed to develop LeIFN-Con, i.e., to develop the [Arg²², Ala⁷⁶, Asp⁷⁸, Glu⁷⁹, Tyr⁸⁶, Tyr⁹⁰, Leu⁹⁶, Thr¹⁵⁶, Asn¹⁵⁷, Leu¹⁵⁸] analog of IFN- α F. The illustrated top strand sequence includes, wherever possible, codons noted to the subject of preferential expression in *E. coli*. The sequence also includes bases providing recognition sites for SalI, HindIII, and BstE2 at positions intermediate the sequence and for XbaI and BamHI at its ends. The latter sites are selected for use in incorporation of the sequence in a pBR322 vector, as was the case with the sequence developed for IFN- α F and its analogs.

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-1 1                                10
Met-Cys-Asp-Leu-Pro-Gln-Thr-His-Ser-Leu-Gly-Asn-Arg-Arg-
ATG TGT GAT TTA CCT CAA ACT CAT TCT CTT GGT AAC CGT CCG
                                20
Ala-Leu-Ile-Leu-Leu-Ala-Gln-Met-Arg-Arg-Ile-Ser-Pro-Phe-
AGA CTG ATT CTG CTG GCA CAG ATG CGT CGT ATT TCC CCG TTT
                                30
Ser-Cys-Leu-Lys-Asp-Arg-His-Asp-Phe-Gly-Phe-Pro-Gln-Glu-
AGC TGC CTO AAA GAC CGT CAC GAC TTC GGC TTT CCG CAA GAA
                                40
Glu-Phe-Asp-Gly-Asn-Gln-Phe-Gln-Lys-Ala-Gln-Ala-Ile-Ser-
GAG TTC GAT GGC AAC CAA TTC CAG AAA GCT CAG GCA ATC TCT
                                50
Val-Leu-His-Glu-Met-Ile-Gln-Gln-Thr-Phe-Asn-Leu-Phe-Ser-
GTA CTG CAC GAA ATG ATC CAA CAG ACC TTC AAC CTO TTT TCC
                                60
Thr-Lys-Asp-Ser-Ser-Ala-Ala-Trp-Asp-Glu-Ser-Leu-Leu-Glu-
ACT AAA GAC AGC TCT GCT GCT TCG GAC GAA AGC TTG CTO GAG
                                70
Lys-Phe-Tyr-Thr-Glu-Leu-Tyr-Gln-Gln-Leu-Asn-Asp-Leu-Glu-
AAG TTC TAC ACT GAA CTO TAT CAG CAG CTG AAC GAC CTG GAA
                                80
Ala-Cys-Val-Ile-Gln-Glu-Val-Gly-Val-Glu-Glu-Thr-Pro-Leu-
GCA TGC GTA ATC CAG GAA GTT GGT GTA GAA GAG ACT CCG CTO
                                90
                                100
                                110

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